



3058 Research Drive
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Analytical Report

PFOA and PFOS Analysis of Deer Muscle and Liver Samples by LC/MS/MS

MPI Report No. L0019346

Testing Laboratory

MPI Research, Inc.
3058 Research Drive
State College, PA 16801

Requester/Project Manager

Dena Haverland
Dalton Utilities
PO BOX 869
Dalton, GA 30722
Phone: 706-529-1010

2010 JAN -6 P 1:44

1 Introduction

Results are reported for the analysis of deer muscle and liver samples received at MPI Research from Dalton Utilities. The MPI Research study number assigned to the project is L0019346. Table I lists the target analytes quantitated for the samples.

Table I. Target Analytes for Quantitation

Compound Name	Acronym
Perfluorooctanoic Acid	C8 Acid or PFOA
Perfluorooctanesulfonate	C8 Sulfonate or PFOS

2 Sample Receipt

Four samples were received from Dena Haverland at Dalton Utilities for this study. The samples were collected on October 02, 2009. The samples arrived on October 06, 2009 via Fedex and were logged in under MPI Research login number L0019346. The shipment was received frozen on dry ice. The samples were stored frozen at approximately -20°C from receipt until analysis. Chain-of-custody information is presented in Attachment A.

3 Methods - Analytical and Preparatory

3.1 Muscle and Liver Sample Preparation

- 3.1.1. Weigh 1 g of muscle or liver sample into a 50 mL disposable centrifuge tube and fortify, if appropriate. Add 0.2 mL of a 100 ng/mL WIS for a final concentration of 1 ng/mL.
- 3.1.2. Add water to the sample for a final volume of 10 mL. Cap tightly.
- 3.1.3. Homogenize sample using a tissuemizer for ~1 minute.
- 3.1.4. Transfer 1 mL of the sample using a disposable pipette into 15 mL disposable centrifuge tubes. Add 5 mL of ACN and shake for ~20 minutes on a wrist action shaker.
- 3.1.5. Centrifuge tubes at ~3000 rpm for ~5 minutes. Carefully decant supernatant into a 50 mL disposable centrifuge tube and add 35 mL of water.
- 3.1.6. Place the unconditioned SPE columns on the vacuum manifold. Condition the SPE columns by passing ~10 mL of methanol through the column followed by ~5 mL of water. The washes may be pulled through the SPE column using vacuum at a flow rate of ~1 drop/sec or may be allowed to pass through the column unaided. Discard all washes. Do not allow the column to dry.
- 3.1.7. Load the sample onto a conditioned SPE column. Discard the eluate. Any analyte residues will be trapped on the SPE column at this point.
- 3.1.8. Elute with 2 mL of methanol. Collect 2 mL of elute into a graduated 15 mL centrifuge tube.

3.2 Sample Analysis by LC/MS/MS

In High Pressure Liquid Chromatography (HPLC), an aliquot of extract is injected and passed through a liquid-phase chromatographic column. Based on the affinity of the analyte for the stationary phase in the column relative to the liquid mobile phase, the analyte is retained for a characteristic amount of time. Following HPLC separation, mass spectrometry provides a rapid and accurate means for analyzing a wide range of organic compounds. Molecules are ionized, fragmented, and detected. The ions characteristic of the compounds are observed and quantitated against external calibration standards.

An HP1100 system interfaced to an Applied Biosystems API 4000 LC/MS/MS was used to analyze the sample extracts for quantitation. A gradient elution through a Phenomenex Luna 3 μ C8(2) Mercury, 20 x 4.0 mm column was used for separation.

The following gradient was performed:

Mobile Phase (A): 2mM Ammonium Acetate in Water
Mobile Phase (B): Methanol

Time	%A	%B
0.0	90	10
0.5	90	10
2.0	10	90
5.0	10	90
5.1	0	100
6.0	0	100
6.1	90	10
10.0	90	10

The following parameters were used for operation of the mass spectrometer:

Parameter	Setting
Ionization Mode	Electrospray
Polarity	Negative
Transitions Monitored	413→369 (PFOA) 499→80 (PFOS) 415→370 (Internal Std. ¹³ C PFOA (m+2)) 503→80 (Internal Std. ¹³ C PFOS (m+4))
Gas Temperature	450°C

4 Analysis by LCMSMS

4.1 Calibration

For the muscle and liver sample analysis, a 6-point calibration curve was analyzed throughout the analytical sequence for PFOA and PFOS. The calibration points were prepared at 0.1, 0.2, 0.5, 1.0, 2.0, 5.0 ng/mL (ppb) each containing 1.0 ng/mL ¹³C-PFOA (m+2) and ¹³C-PFOS (m+4).

The ratio of the analyte concentration to the IS concentration versus the ratio of the analyte instrument response (area) to the IS response (area) was plotted for each point. Using linear regression with 1/x weighting, the slope, y-intercept and coefficient of determination (r^2) were determined. A calibration curve is acceptable if $r^2 \geq 0.985$.

For the results reported here, calibration criteria were met. The calibration curves are included in the raw data in Attachment C.

4.2 Laboratory Control Spikes

Laboratory control spikes in the analytical set were prepared during each extraction set by adding a known concentration of the analyte to deer muscle and liver controls. Laboratory control spikes are used to assess method accuracy. The laboratory control spikes must show recoveries between 70-130% or the data is rejected. For the results reported here, the laboratory control spikes were within the acceptable range. Laboratory control spike recoveries are given in Attachment B.

4.3 Matrix Spikes

A matrix spike was prepared for each sample by adding a known concentration of the target analyte to a sample. Matrix spikes are used to assess method accuracy in the matrix. The matrix spikes should show recoveries between 70-130%. For the results reported here, the matrix spike was within the acceptable range with the exceptions of:

L19346-4 (Deer # 7 3.5 yr male liver) Spk C at 2000 ng/g for PFOS, which gave low recovery of 59%.

4.4 Laboratory Duplicates

Each sample was prepared in duplicate and analyzed. Duplicate results are given along with the sample results in Attachment B.

5 Data Summary

Please see Attachment B for a detailed listing of the analytical results. For the muscle and liver samples the results are reported in parts per billion (ng/g) on an as-received basis.

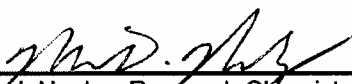
6 Data/Sample Retention

Samples are disposed of 60 days after the report is issued unless otherwise specified by the project manager. All electronic data is archived on retrievable media and hard copy reports are stored in data folders maintained by MPI Research. Hardcopy data is stored for a minimum of five years. The client will be notified 30 days prior to the disposal of hardcopy data.

7 Attachments

- 7.1 Attachment A: Chain of Custody
- 7.2 Attachment B: Analytical Results
- 7.3 Attachment C: Raw Analytical Data for Water


8 Signatures



Mark Neeley, Research Chemist Associate II

11-6-09

Date



Robert Zhu, Manager, Analytical

11/9/09

Date



A





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54943 North Main Street
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3058 Research Drive
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(814) 231-1580 Fax

Login

Login Group: L0019346

Login #: 19460
Project: P0005196
Company Name: Dalton Utilities
Submitted By: Dena Haverland
Login Type: Immediate Receipt of Samples
Started: True
Date Start: 10/27/2009
Due Date: 11/06/2009
Login Initiated: 10/27/2009
Received By: Ammerman, Mark
Spread Sample:
Label:
MPI SD/PI: Zhu, Xiang
Project Title/Type: PFOA and PFOS Analysis of Animal Muscle and Liver by LC/MS/MS / ROUTINE
Login Notes:

Conform COC Sample: True
Conform COC: True
Conform Sample: True
Conform Request: True

Packages / Containers

Package	Carton	Date / Condition	Shipper / ID	Temp. Control/Temp.	Direction / Handled By
K0022042		Received Date: 10/27/09 10:25 Package & Contents Uncompromised	FEDEX 8694 2057 8178	Dry Ice -79.2	RECEIVED Ammerman, Mark

Container #	Gross Weight	pH	Container Type	Preservative	Mfg. Lot	Mfg. ID
C0457624	218.20 g		1/2 gallon ziplock bag	NONE		
C0457625	278.00 g		1/2 gallon ziplock bag	NONE		
C0457626	324.10 g		1 gallon ziploc bag	NONE		
C0457627	994.10 g		1 gallon ziploc bag	NONE		

Samples

Sample ID	Container	Matrix	System	System Matrix	Sample	Date Sampled	Date Due
L0019346-0001	C0457624	SOLID	Deer	Tissue	Deer #6 0.5 yr female-muscle	10/02/2009	11/06/2009
L0019346-0002	C0457626	SOLID	Deer	Liver	Deer #6 0.5 yr female-liver	10/02/2009	11/06/2009
L0019346-0003	C0457625	SOLID	Deer	Tissue	Deer #7 3.5 yr male-muscle	10/02/2009	11/06/2009
L0019346-0004	C0457627	SOLID	Deer	Liver	Deer #7 3.5 yr male-liver	10/02/2009	11/06/2009



Login

Login Reviewed By:



Date/Time:

10/28/09 1440



MPI

RESEARCH

MPI Research Contact: Daniel Wright

Send Report To:

Company: Dalton Utilities
 Address: 1200 VD Parrott JR Parkway, PO Box 869
 City, State, ZIP: Dalton, GA 30722-0869
 Attention: Dena Haverland
 Phone #: 706-529-1010
 Fax #: 706-529-1271
 Email: dhaverland@dutil.com
 Study/Job #: _____
 Signature/Date: _____
 Printed Name: _____

Sample Submittal

Please fax this form before sending samples.

Please send samples to shipping and receiving:
 3048 Research Drive, State College, PA 16801
 T: (814) 272-1039 • F: (814) 272-1019

Turnaround time (TAT) requirements:

Results Due Date: 30 days
 Preliminary Results Format: Verbal Email Fax
 Report Due Date: 30 days

Storage Conditions

Room temperature
 Refrigerator
☒ Freezer
 Ultra Low freezer
 Desiccated
 Lighting required

Stability (°C/%RH): _____

Stability time period: _____

Safety Information

Special handling: _____
 MSDS attached
 Controlled substance: _____
 HAZARDS: _____
 Please fill in the diamond HMIS/NFPA
 (0 4) if appropriate

Client ID# Description	Lot/ Control #	Amt. Sent/ Weight	# of Bottles	Matrix	Date & Time	Tests Requested
1 <u>Deer #6</u> <u>0.5 yr Female-Serum</u>		<u>10µl</u>	<u>10</u>	<u>deer</u>	<u>10/2/09 1:08AM</u>	<u>PFOA/PFOS</u>
2 <u>Deer #6</u> <u>0.5 yr Female-muscle</u>		<u>repackaged</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:28AM</u>	<u>PFOA/PFOS</u>
3 <u>Deer #6</u> <u>0.5 yr Female-Liver</u>		<u>Whole</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:36AM</u>	<u>PFOA/PFOS</u>
4 <u>Deer #7</u> <u>3.5 yr Male-Serum</u>		<u>10µl</u>	<u>10</u>	<u>deer</u>	<u>10/2/09 1:45AM</u>	<u>PFOA/PFOS</u>
5 <u>Deer #7</u> <u>3.5 yr Male-muscle</u>		<u>repackaged</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:45AM</u>	<u>PFOA/PFOS</u>
6 <u>Deer #7</u> <u>3.5 yr Male-Liver</u>		<u>Whole</u>	<u>1 bag</u>	<u>deer</u>	<u>10/2/09 2:48AM</u>	<u>PFOA/PFOS</u>
7						
8						
9						
10						

PO # _____

Relinquished by: Dan Time: 10/5/09 6:30AM Received by: Dan Date: 10/6/09 10:25

Notes

THIS IS AN EXACT COPY OF
 THE ORIGINAL DOCUMENT

BY MLA DATE 10/27/09



TEMPORARY SAMPLE STORAGE FORM

To be completed during ExyLIMS Login

Project #: 15196

Login #: L 14346

Initials / Date: MC 10/27/09

One form to be completed for each package

Date / Time Received: 10/26/09 1025

Received By: Mark Ammerman

Shipper: FedEx

Shipper Package ID: 8694 2057 8178

Temperature (deg C) / Thermometer ID: -79.2 / D0001775

Temperature Control Method: dry ice - active

Temporary Storage Location: Walkin Freezer 11

Condition of sample(s):

- ☒ Good – Package and contents uncompromised
☐ Fair – Package damaged / contents uncompromised
☐ Poor – Package and contents compromised

Notes:

FedEx Express **US Airbill**

FedEx Tracking Number **8694 2057 8178**

1 From **10/5/99**

Sender's Name **Daniel K. Hargrave** Phone **204-246-2009**

Company **USDA Wildlife Service**

Address **200 Phoenix Road**

City **Alhambra** State **CA** ZIP **91802**

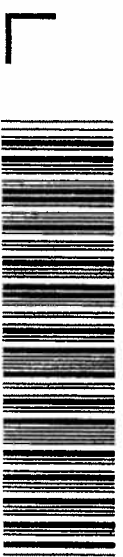
2 Your Internal Billing Reference

3 To Recipient's Name **Daniel K. Hargrave** Phone **204-246-2009**

Company **MPT Research Labs**

Address **3044 Rockwood Drive**

City **Little Rock** State **AR** ZIP **72201**



8694 2057 8178

Recipient's Copy

4a Express Package Service

☒ FedEx Priority Overnight ☐ FedEx Standard Overnight

☐ FedEx 2Day ☐ FedEx Express Saver

4b Express Freight Service

☐ FedEx 1Day Freight ☐ FedEx 2Day Freight

5 Packaging

☐ FedEx ☐ FedEx Pay* ☐ FedEx Box ☐ FedEx Tube ☒ Other

6 Special Handling

☐ SATURDAY Delivery ☐ HOLD Weekday ☐ HOLD Saturday

☒ No ☐ Yes ☐ Dry Ice ☐ Cargo Aircraft Only

7 Payment

☒ Sender ☐ Recipient ☐ Third Party ☐ Credit Card ☐ Cash/Check

Total Packages **1** Total Weight **1.00** Total Declared Value **\$.00**

8 Residential Delivery Signature Options

☒ No Signature Required ☐ Direct Signature ☐ Indirect Signature

520



B





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Analytical Report

Summary of Fluorochemical Residues in Muscle Samples

Sample ID	PFOA	PFOS
	Perfluorooctanoic Acid	Perfluorooctanesulfonate
	Analyte Found (ng/g, ppb)	Analyte Found (ng/g, ppb)
Deer # 7 3.5 yr male-muscle	ND	88.4
Deer # 7 3.5 yr male-muscle*	ND	92.2
Deer # 6 0.5 yr female-muscle	ND	13.4
Deer # 6 0.5 yr female-muscle*	ND	11.9

*Laboratory Duplicate

ND = Not detected = Response is below the LOD of 1.0 ng/g (ppb).

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g (ppb).



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Analytical Report

Recovery Summary of Fluorochemical Residues in Muscle Samples

Sample Description	Amount Spiked (ng/g)	Amt Found In Sample (ng/g)	PFOA		Amt Found In Sample (ng/g)	PFOS	
			Amount Recovered (ng/g)	Recovery (%)		Amount Recovered (ng/g)	Recovery (%)
LCS A (Data set 110409A) 10 ng/g	10	ND	9.37	94	ND	8.78	88
LCS B (Data set 110409A) 50 ng/g	50	ND	47.1	94	ND	45.2	90
Deer # 7 3.5 yr male-muscle (L19346-3 Spk C, 50 ng/g Lab Spike)	50	ND	48.5	97	88	128	79
Deer # 6 0.5 yr female-muscle (L19346-1 Spk D, 50 ng/g Lab Spike)	50	ND	46.4	93	13.4	58.0	89

ND = Not detected = Response is below the LOD of 1.0 ng/g.

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g.



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Summary of Fluorochemical Residues in Liver Samples

Sample ID	PFOA Perfluorooctanoic Acid	PFOS Perfluorooctanesulfonate
	Analyte Found (ng/g, ppb)	Analyte Found (ng/g, ppb)
Deer # 7 3.5 yr male-liver	ND	2150^
Deer # 7 3.5 yr male-liver*	ND	2160^
Deer # 6 0.5 yr female-liver	ND	604
Deer # 6 0.5 yr female-liver*	ND	574

*Laboratory Duplicate

ND = Not detected = Response is below the LOD of 1.0 ng/g (ppb).

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g (ppb).

^ The corresponding Matrix Spike recovery was outside the acceptance criteria of 70-130%, therefore the sample values should be considered an estimate.



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Recovery Summary of Fluorochemical Residues in Liver Samples

Sample Description	Amount Spiked (ng/g)	Amt Found in Sample (ng/g)	PFOA		Amt Found in Sample (ng/g)	PFOS	
			Amount Recovered (ng/g)	Recovery (%)		Amount Recovered (ng/g)	Recovery (%)
LCS A (Data set 110409B) 10 ng/g	10	ND	10.0	100	16.5	26.3	98
LCS B (Data set 110409B) 50 ng/g	50	ND	50.3	101	16.5	61.6	90
LCS A (Data set 110509B) 10 ng/g	10	N/A	N/A	N/A	18.6	28.8	102
LCS B (Data set 110509B) 50 ng/g	50	N/A	N/A	N/A	18.6	67.9	99
Deer # 7 3.5 yr male-liver (L19346-4 Spk C, 50 ng/g Lab Spike)	50	ND	52.4	105	2150	**	**
Deer # 6 3.5 yr female-liver (L19346-2 Spk D, 50 ng/g Lab Spike)	50	ND	50.4	101	604	**	**
Deer # 7 3.5 yr male-liver (L19346-4 Spk C, 2000 ng/g Lab Spike)	2000	N/A	N/A	N/A	2150	3330	59*
Deer # 6 3.5 yr female-liver (L19346-2 Spk D, 250 ng/g Lab Spike)	250	N/A	N/A	N/A	604	817	85

ND = Not detected = Response is below the LOD of 1.0 ng/g.

NQ = Not quantifiable = Response is between the LOD and the LOQ of 10 ng/g.

* The Matrix Spike recovery was outside the acceptance criteria of 70-130%, therefore the sample values should be considered an estimate.

** The endogenous level of PFOS in the sample significantly exceeds the spiking level, therefore an accurate recovery cannot be calculated.